Detection of Viral Vector Sequences in Animal Excretions

Dawn P. Wooley, Ph.D., SM(NRCM), RBP, CBSP

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Outline

- Previous Study
- Current Study
 - Vector Systems
 - QPCR Assay
 - Virus Stocks
 - Animal Experiments
 - Results
 - Conclusion
- Future Direction

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Original Research

Assessment of Hazard Risk Associated with the Intravenous Use of Viral Vectors in Rodents

Jon D Reuter,12,* Xiaoqun Fang,3 Christina S Ly,3 Karen K Suter,1 and Daniel Gibbs3

¹Animal Resources Department, ²Gene Expression Laboratory, and ³GT3 Core Facility, The Salk Institute for Biological Studies, La Jolla, California. ^{*}Corresponding author. Email: reuter@salk.edu

"...we conclude that commonly used replication-deficient viral vectors pose minimal risk by 72 hours after inoculation....Level 1 safety measures may be sufficient after cage changing and biosafety evaluation."

Previous Shedding Study

- Mice, 8-wk-old females
 - Outbred strain
 - Immune deficient
- Vector Viruses
 - Lenti and Adeno
 - EGFP transgene driven by CMV promoter
 - ~10⁹ IU/mouse
- Urine and Feces
 - Collected as excreted
 - Time points -1, 1, 3, 7

Previous Shedding Study (continued)

- Isolated nucleic acids
- Performed quantitative PCR
 - Primers against the transgene
 - No internal control
- Positive controls
 - Spiked whole blood with vector virus
 - Limit of detection was 200
 - Used as control for urine and fecal samples
- Result
 - No shedding detected for Lenti and Adeno



High Pure Viral Nucleic Acid Large Volume Kit, Roche

AdEasy[™] Adenoviral Vector System



ViraPower[™] Lentiviral Expression System (HIV)



Multiplex PCR





<u>Ours</u>

Quality Control (QC) Sequences



Virus Stock Characterization

Stock	Infectious Titer (pfu/ml or cfu/ml)	Total No. of Virus Particles (vg/ml)	Total Particle: Infectious Particle Ratio	p24 Protein Content (µg/ml)
Adeno	4 x 10 ⁸	2 x 10 ⁹	5	N/A
Lenti	1 x 10 ⁷	3 x 10 ¹¹	3 x 10 ⁴	13.3 ^{<i>a</i>}

N/A = Not Applicable ^aEquivalent to 2.7 x 10¹¹ particles/ml

Experimental Scheme



Mouse Experimental Setup



Excretion Outputs from Infected Mice

Adeno Vector in Mice (Summer months)

- Urine output was 1.9 ml/day/mouse (0.75-3.8)
- Fecal output was 1.7 g/day/mouse (0.46-3.8)

Lenti Vector (Winter months)

- Urine output was 0.9 ml/day/mouse (0.11-2.1)
- Fecal output was 1.2 g/day/mouse (0.29-2.1)

Daily Fecal Excretions





2.298 g / 96 pellets = 24 mg per pellet

DNA Isolations

QIAamp MinElute Virus Spin Kit

QIAamp DNA Stool Mini Kit



QPCR Experiment with Plasmid-Spiked Urine for Adenoviral Vector



Note: Sample was spiked just after addition of lysis buffer and carrier RNA using Qiagen MinElute Kit

QPCR Experiment with <u>Virus</u>-Spiked Urine for Adenoviral Vector



Conclusions

- Shedding studies are technically challenging, not a trivial undertaking
- Be cautious with interpreting negative results
- Robust positive controls must be designed
- Limits of detection must be accurately established
- Sample size must be considered
- Virus particles must be kept intact as long as possible
- Viral lysis should be performed as far downstream as possible.

Ongoing and Future Studies

- Develop a more sensitive way of processing the current samples
- Higher doses of vector viruses
- Different routes of administration
- Different Lentivirus and Adenovirus vector constructs
- Armored RNA and DNA controls

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"A pessimist is simply an optimist in full possession of the facts."

Edward Abbey