Validation and Utility of a Bleach Chemical Effluent Decontamination System (CEDS) for

Biocontainment Laboratories

Dr. David Harbourt November 20, 2019

Celebrating 50 Years

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Outline

- Laboratory testing of bleach efficacy
- Process for Chemical Effluent Decontamination System (CEDS)
 Validation
- Validation Results
- Utility of CEDS for Biocontainment Laboratories
- Discussion

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Laboratory Testing

- Initial proof of concept work performed using dialysis tubing in bleach solution
- Tested at various concentrations/contact times and with and without organics
- Relied on *B. thuringiensis (Bt)* spores for efficacy



Laboratory Testing Procedure

Prepare the bleach solution and add 5% FBS Prepare spore inoculum of Bt (in Tween solution)

Heat spore _____ suspension at 65C for 30 min Prepare dialysis tubing by prewetting in PBS With sterile scissors Load 2 ml of spore suspension cut the dialysis tubing at the end with the alligator clip

Suspend the spore packets for the dilution of spores on blood agar

After desired contact time, retrieve the spore packet and suspend over sterile flask (1 liter flask with 500 ml Heart Infusion Broth -HIB)

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Laboratory Testing Procedure (continued)

Incubate flasks at 37C with shaking at 150 rpm

100uL spore culture+2mL bleach solution added to flask to incubate at 37C with shaking at 150 rpm



Parameter tested ¹	total spore	time (h)	Interday variation	# total samples	pass	fail	failure rate ²
11,400 ppm	4x10 ⁶	2	no	3	3	0	0
11,400 ppm + 1% FBS	4x10 ⁶	2	no	3	3	0	0
5,700 ppm + 5% FBS	4.5-7.8 x10 ⁶	2	yes	9	9 ³	0	0
5,700 ppm + 5% FBS	3.6-4.5 x10 ⁶	1	yes	6	5	1	17%
5,700 ppm + 5% FBS	3.6-4.5 x10 ⁶	0.5	yes	6	3	3	50%
3,800 ppm + %5 FBS	3.6x10 ⁶	20	no	3	3 4	0	0%
3,800 ppm + %5 FBS	3.6x10 ⁶	2	no	3	2	1	33%
3,800 ppm + %5 FBS	3.6x10 ⁶	1	no	3	0	3	100%
3,800 ppm + %5 FBS	3.6x10 ⁶	0.5	no	3	0	3	100%

¹ppm is based upon % free chlorine per each bleach source

²failure rates reflective of past work at USAMRIID and published literature

³expt pending: 2 independent trials running, sterile after 30 h; so if sterility maintained N=9 samples,

3 distinct bottles of bleach

⁴passed through day 4 final results pending

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EDS Validation Process



Sample	Free Chlorine (ppm)	Free Chlorine (ppm)	Free Chlorine (ppm) 120 min	
	0 min	15 min		
Run A				
High	6600	6600	6400	
Low	6800	6400	6400	
Run B				
High	6600	6600	6400	
Low	6600	6600	6600	
Sample	Free Chlorine (ppm) 0 min	Free Chlorine (ppm) 15 min	Free Chlorine (ppm) 120 min	
Run C				
High	6400	6400	6400	
Low	6400	6400	6400	
Run D				
High	6200	6400	6600	
Low	6200	6400	6600	
Run E				
High	6400	6400	6600	
Low	6400	6400	6400	
Run F				
High	6400	6200	6400	
Low	6600	6400	6200	
Run G				
High	6400	6800	6400	
Low	6400	6600	6200	
Run H				
High	6400	6600	6600	
Low	6200	6400	6400	
Run I				
High	6400	6400	6600	
Low	6400	6600	6400	
Run J				
High	6400	6400	6200	
Low	6600	6600	6600	

Table 1. Free chlorine concentrations for consistency run testing/pre-validation

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Biological Challenge Procedure

Gather PPE for CEDS entry

Prepare spore inoculum of Bt (in Tween solution)

Heat spore suspension at 65C for 30 min Prepare dialysis Load 2 ml of spore tubing by prewetting in sterile dialysis tube PBS

Enter CEDS in PPE to load fishing line with Tyvek coupons and spore suspensions in CEDS tank

Perform serial dilution of spores on blood agar Place metal coupons containing 10⁶ spores inside Tyvek envelopes on fishing line weighted down with washers Place samples in beaker with buffer solution for transport

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Biological Challenge Procedure Continued

After 2 hours, retrieve the fishing line with Tyvek coupons and spore suspensions

Tyvek coupons to microbiology core lab

Results read and reported

Spore suspensions to With sterile scissors (performed with 2 lab people) cut the dialysis tubing at the end with the alligator clip Sample and positive controls are incubated and analyzed in Celebr same manner and laboratory testing procedures

Sample	rree Chiorine (ppin)		
Run #1			
High	6400		
Low	6600		
Run #2			
High	6400		
Low	6400		
Run #3			
High	6800		
Low	6600		
Run #4			
High	6400		
Low	6400		
Run #5			
Sample	Free Chlorine (ppm)		
Run #6			
High	6600		
Low	6400		

Biological Challenge Results: Tyvek Coupons

C	Broth Culture	Agar culture	
Sample	(72 hours)	(72 hours)	
Run #1-positive ctl	Positive	Positive	
High	Negative	Negative	
Middle	Negative	Negative	
Low	Negative	Negative	
Run #2-positive ctl	Positive	Positive	
High	Negative*	Negative*	
Middle	Negative	Negative	
Low	Negative	Negative	
Run #3-positive ctl	Positive	Positive	
High	Negative	Negative	
Middle	Negative	Negative	
Low	Negative	Negative	
Run #4-positive ctl	Positive	Positive	
High	Negative	Negative	
Middle	Negative	Negative	
Low	Negative	Negative	
Run #5-positive ctl	Positive	Positive	
High	Negative	Negative	
Middle	Negative	Negative	
Low	No recovery	No recovery	
Run #6-positive ctl	Positive	Positive	
High	Positive**	Positive**	
Middle	Negative	Negative	
Low	Positive**	Positive**	

*Indicates growth was seen in broth and agar but colonies were gram negative bacteria consistent with environmental contamination **Indicated growth was seen in broth and agar due to insufficient contact between bleach and spore coupons. It is likely that there was inadequate space for bleach solution to penetrate the Tyvek envelope preventing inactivation. This explanation is corroborated by the negative results from the USAMRIID-prepared spore packets from the same runs.

Biological Challenge Results: Spore Suspensions

S	Broth Culture	Optical Density	Agar culture	
Sampie	(7 day)	(OD ₆₂₀)	(7 day)	
Run #1-positive ctl	Positive	0.78	Positive	
High	Negative	0	Negative	
Middle	Negative*	0.36*	Negative*	
Low	Negative	0	Negative	
Run #2-positive ctl	Positive	0.85	Positive	
High	Negative	0	Negative	
Middle	Negative	0	Negative	
Low	Negative	0	Negative	
Run #3-positive ctl	Positive	0.78	Positive	
High	Negative	0	Negative	
Middle	Negative	0	Negative	
Low	Negative	0	Negative	
Run #4-positive ctl	Positive	0.96	Positive	
High	Negative	0	Negative	
Middle	Negative	0	Negative	
Low	Negative	0	Negative	
Run #5-positive ctl	Positive	0.82	Positive	
High	Negative	0	Negative	
Middle	Negative	0	Negative	
Low	Negative	0	Negative	
Run #6-positive ctl	Positive	0.94	Positive	
High	Negative	0	Negative	
Middle	Negative	0	Negative	
Low	Negative	0	Negative	

*Culture was negative for *B*. *thuringiensis*, but was a pure culture of *B*. *atrophaeus*

Additional Considerations for CEDS application in **Biocontainment Labs**

$Na_{2}S_{2}O_{5} + 2HOCI + 2H_{2}O_{5}$ → 2NaHSO₄ + 2 HCl

- Discharge costs to neutralize or disposal of chemically treated effluent
- Additional PPE considerations during operations
- Difficulty of monitoring chemical concentrations
- Efficacy in presence of high organics
- CEDS units do not function properly unless system design permits complete drainage at end of each cycle
- Chemical procurement
- Transport of chemicals between storage and CEDS
- Extensive laboratory testing needed for validation

Types of Chemical Effluent Decontamination Systems

	Bleach	Chlorine Dioxide (CD)	Peroxyacetic Acid (PAA)
Affected by organics	Yes (trihalomethane generation)	No	No
Delivery	Concentrated liquid	Solid, liquid (through acid and sodium chlorite) or gas	Concentrated liquid
Neutralization required	Yes (both pH and dechlorination)	Yes (dechlorination only)	Yes (both PAA and pH)
Efficacy	6000ppm for 2h	1000ppm for 3h	1500ppm for 1h (est. by EPA)
Cost	\$5/gal in bulk for 12.5 % bleach	\$5/gal for sodium chlorite	

Discussion

- We have validated a bleach based CEDS using a minimum concentration of 6200ppm free chlorine and a two hour contact time
- Bleach procurement and disposal considerations make it very difficult to operate a bleach based CEDS over an extended period
- As a general rule, the lowest initial cost solution may not be the right one
- Extensive laboratory work is required both upfront and during any CEDS validation process
- Bleach based CEDS units only have limited utility in biocontainment without significant financial resources

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QUESTIONS??

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